

Metabolite Identification in the Real World



Dr. Axel Paeler



Marieke Teppner

ORGANISED BY:

european
pharmaceutical
review

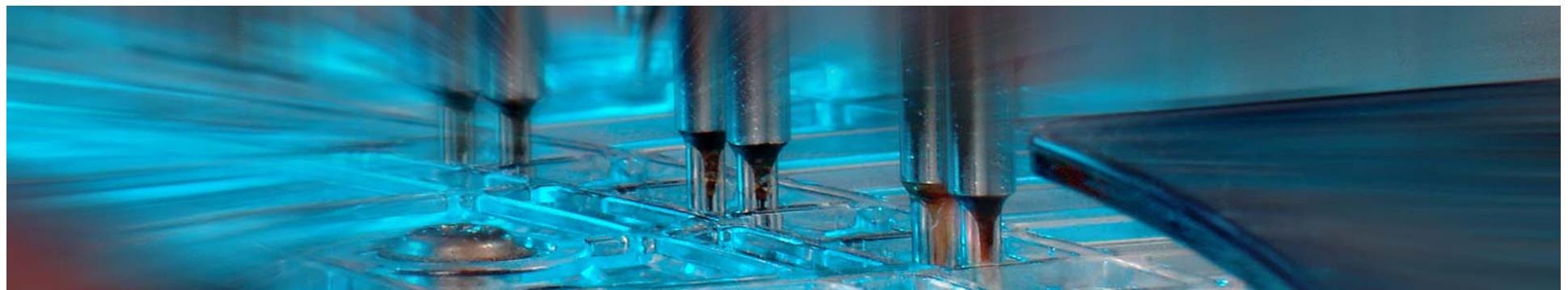
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AB SCIEX

Transforming data into knowledge: Use of HRMS instrumentation in Drug Discovery and Development

Dr. Axel Pähler

DMPK, Non Clinical Safety, pRED, F. Hoffmann-La Roche Ltd.



Drug Discovery & Drug Development

Quantitative metabolite identification workflows



Discovery Research (LI/LO): Find, Quantify? & ID

Hepatic clearance combined with metabolite ID

Parent disappearance kinetics plus relative metabolite formation rates. Can we move from QqQ to HRMS?

Classical metabolite ID study & peak area ratios
(no absolute but relative quantification in tissues/species)



Drug Development: Find, Quantify & ID

Compare human metabolites to animal species

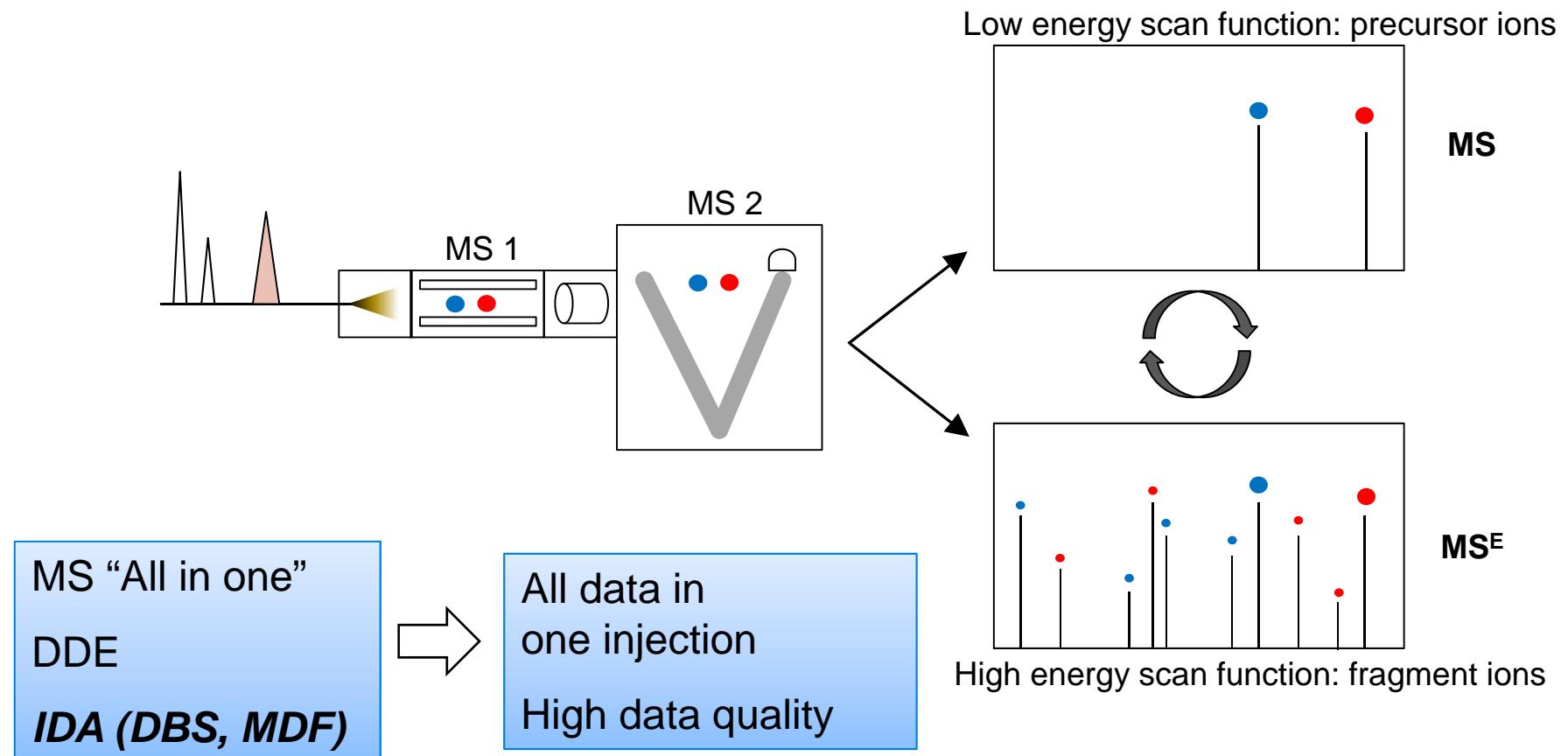
In vitro metabolism (relative peak area ratios or ¹⁴C)

In vivo systemic metabolite exposure (relative peak area ratios, ¹⁴C or authentic standards)

Targeted vs. non Targeted Data Acquisition

MS “*all in one*” or *data dependent* “*true*” MS/MS

- MS^E mode: Two acquisition functions are monitored in parallel
Principally technology independent QqTOF, (q/trap)-FTMS



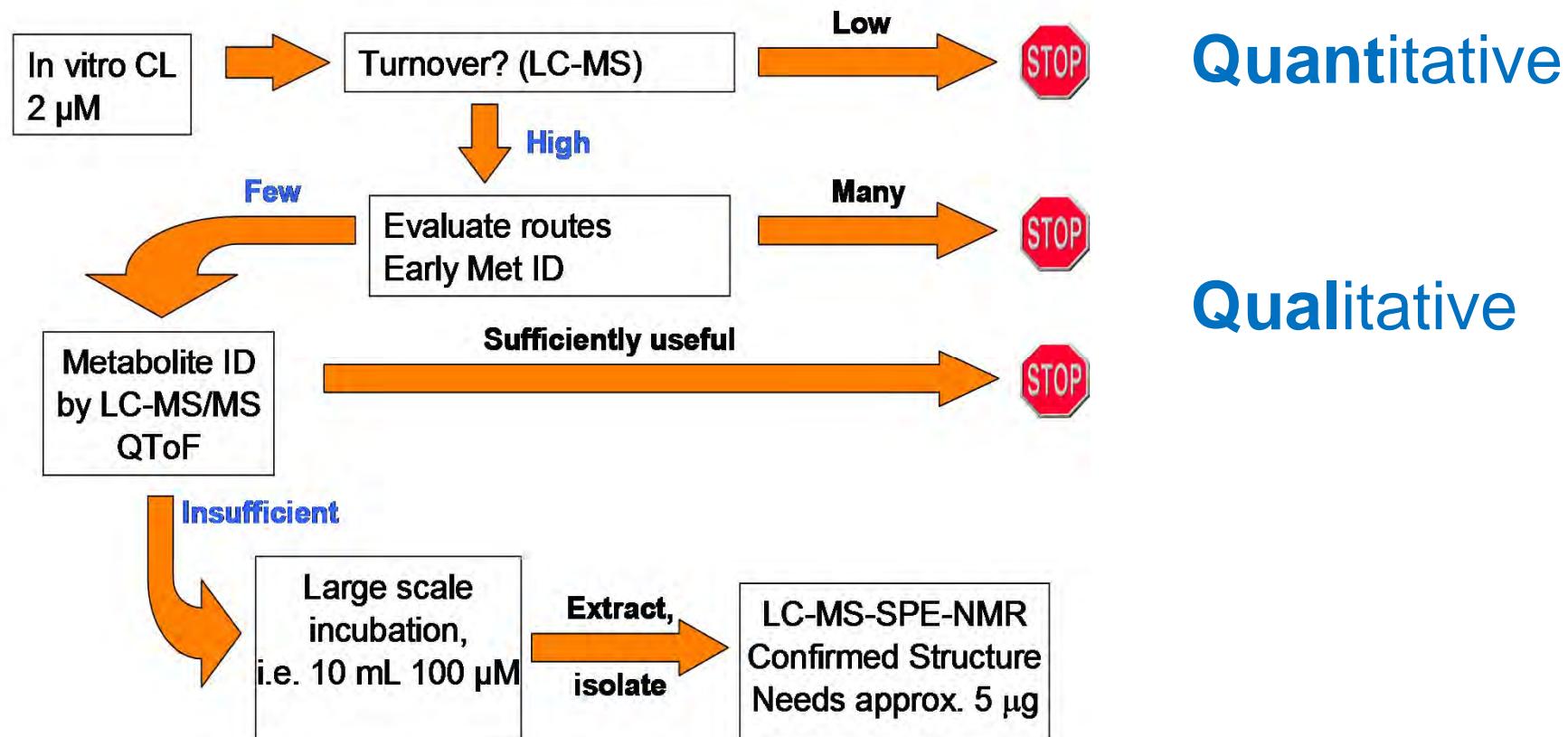
Early Metabolite ID

Shift towards Qual/Quant workflow

SoM identification for high CL compounds

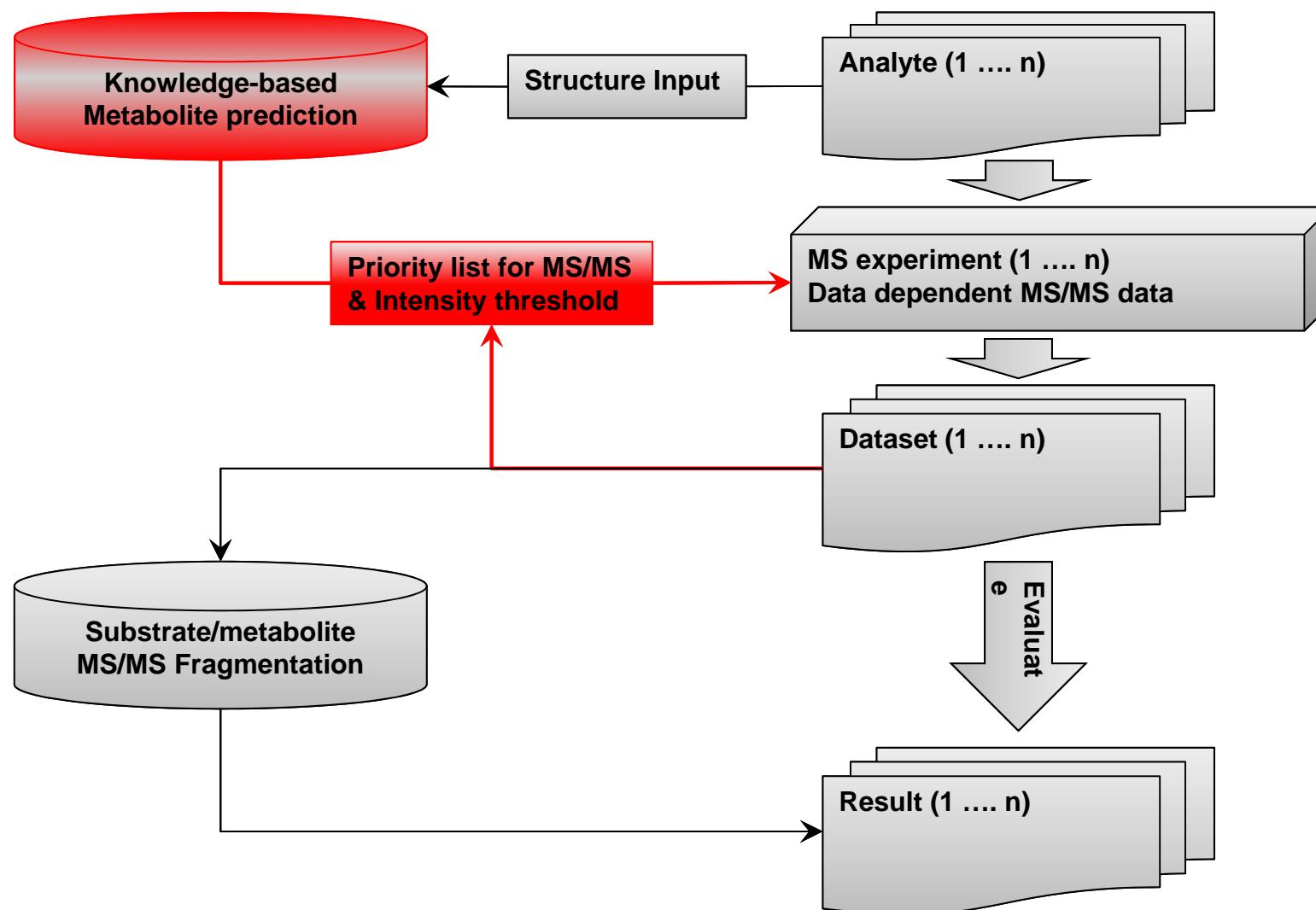
→ Guide rational Drug Design by Medicinal Chemistry

Soft spot ID (top 4 metabolites) → Structure proposal by MS/MS



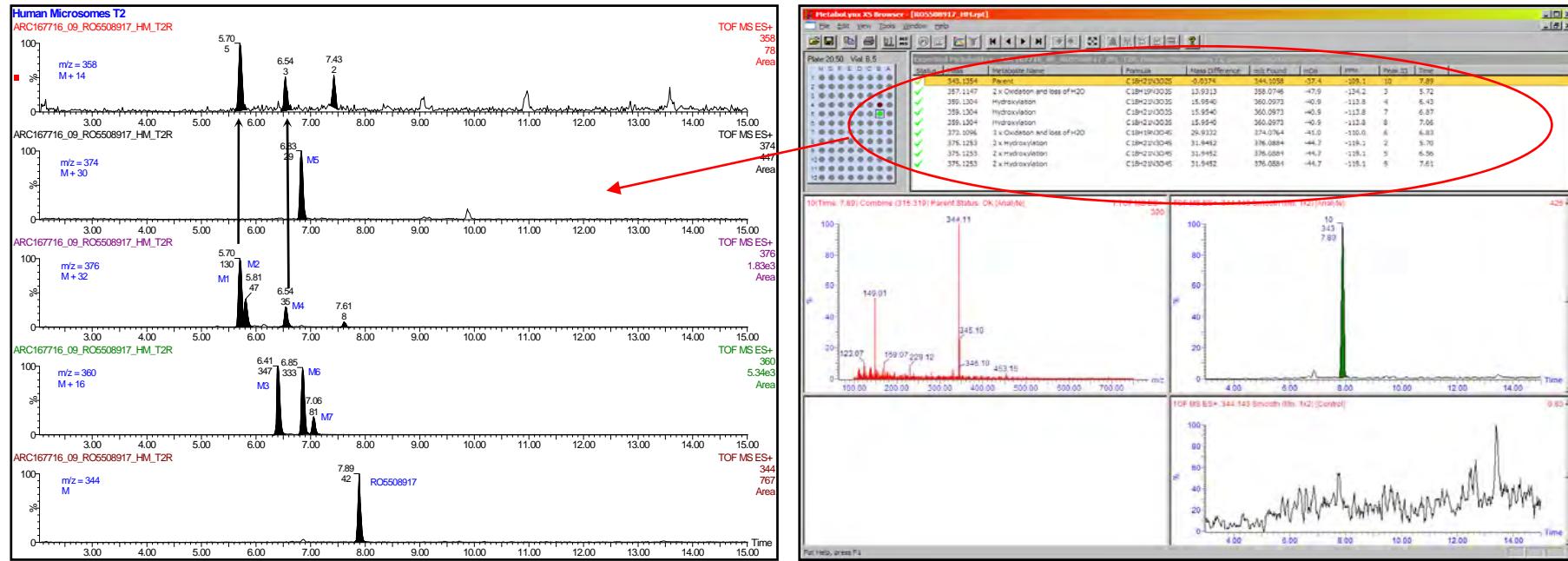
Metabolite Identification: The Past

Qual analysis driven by structure & expert knowledge



From Prediction to Experiment

Data processing and metabolite fishing



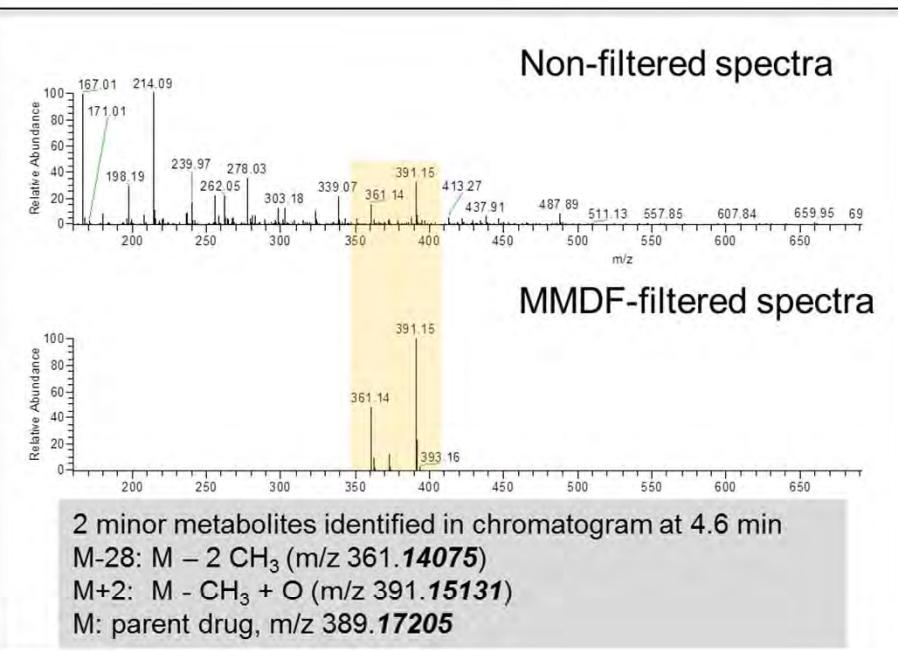
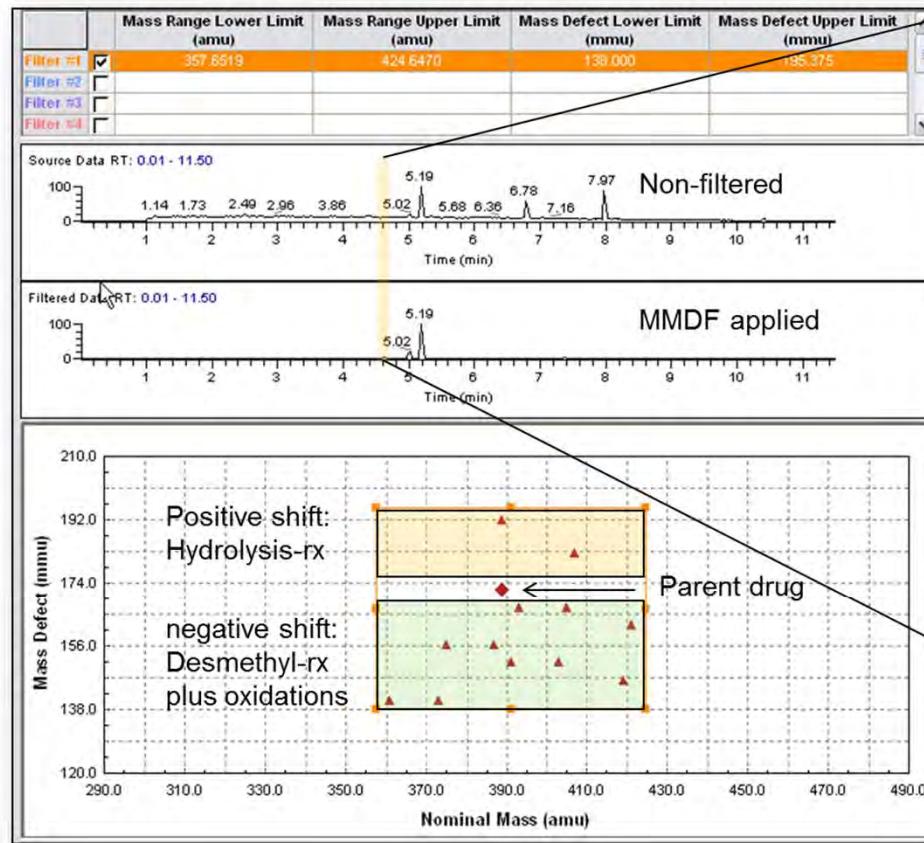
Automated data processing (find expected metabolites)

Search for metabolites based on mass defect filters, isotope clustering and possible de-alkylations: Support detection of possible cleavages and unexpected metabolites

Experienced analysts key for data review and interpretation

Post Acquisition Data Processing

Getting most out of the HRMS data



Mass defect filtering:

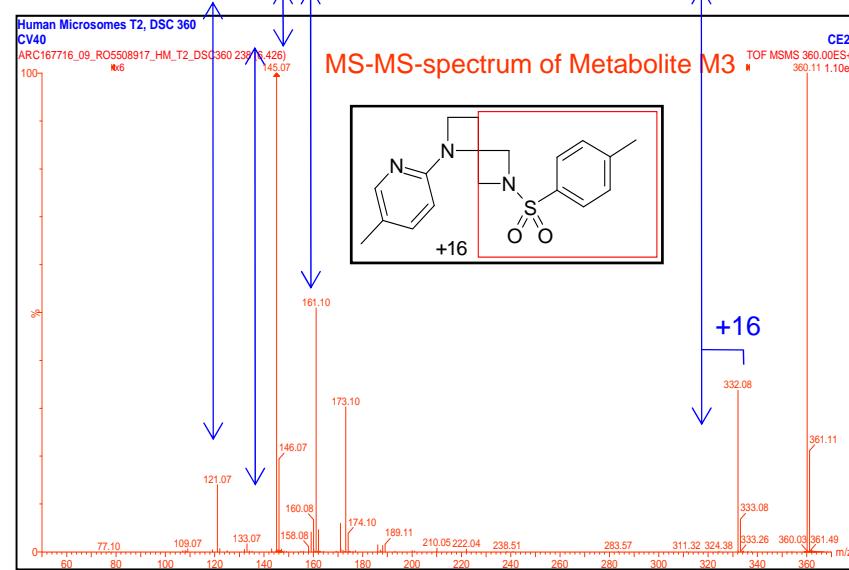
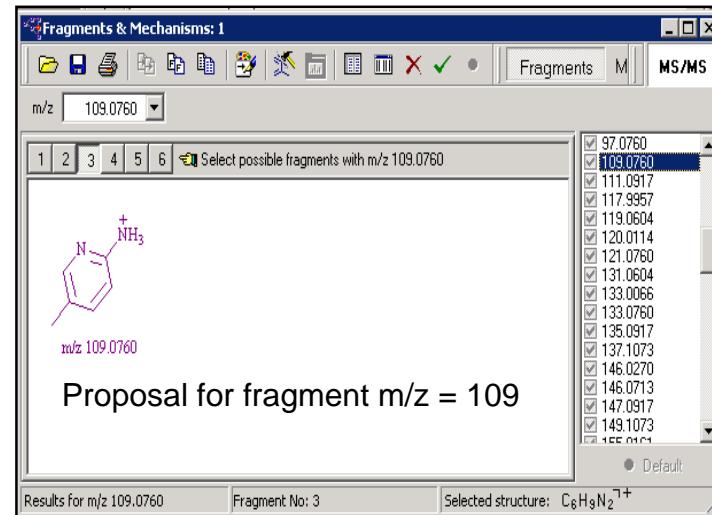
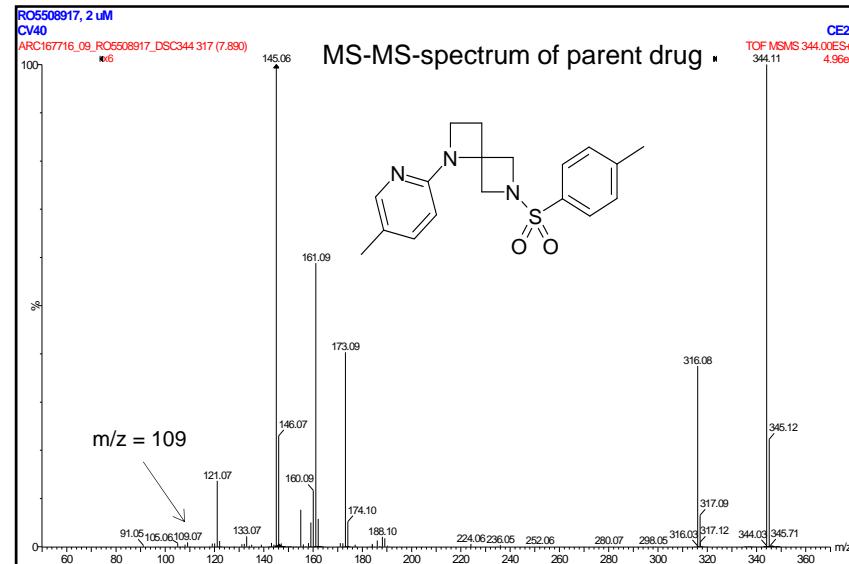
Data reduction

Facilitate finding drug related material

On-the-fly MMDF available on 5600+

From Identification to Structure

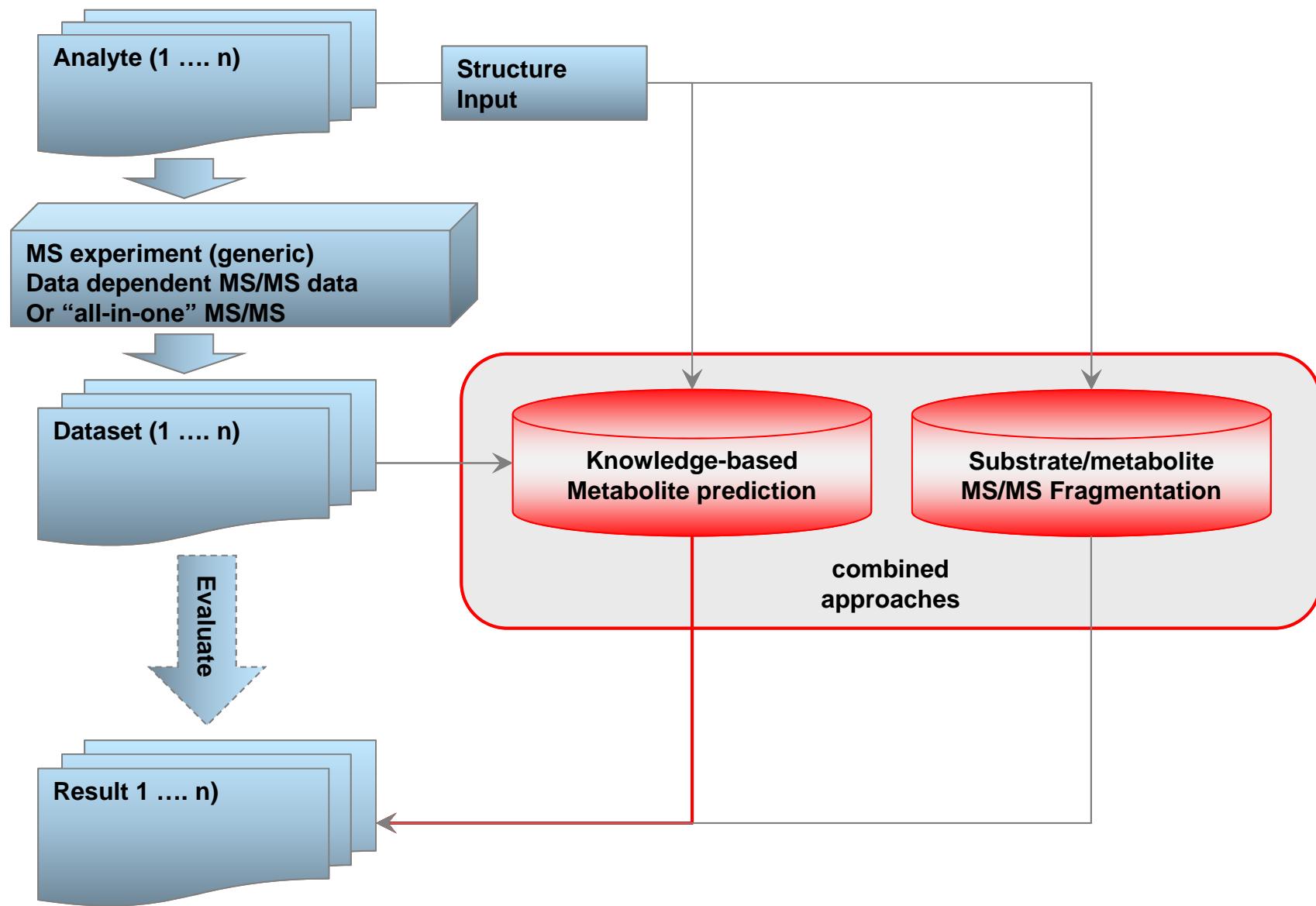
Data processing in Metabolite ID



- Use MS/MS spectra to localize position of biotransformation in the molecule.
- MS/MS spectra interpretation supported by software (based on general fragmentation rules).
- Definitive structure assignment still manual, often Markush-like
- Definitive structure by NMR

Metabolite Identification: The Future

Data driven, untargeted post acquisition analysis



Diversity in Applications

Demand for Versatile LC-MS Instrumentation



High Sensitivity and Speed:

Assays conducted at low physiological drug concentrations
Need to detect relevant metabolites and biomarkers
All information, all the time (full scan MS, MS/MS)

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No significant down time
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Ease of Operation:

Automated setup and calibration routine
Shift from instrumentalist to DMPK specialist as main user
Ready to go, still highly customizable

Software features for DMPK studies



A workflow perspective (A DMPK Scientist's dream)

Vendor independent

Waters (*.raw), Thermo (*.RAW): Ion-Trap/Orbitrap, Agilent (*.d), SCIEX (*.wiff)

Automatic peak detection using multiple algorithms

Targeted/unknown analysis, MDF, fragmentation pattern analysis, noise removal, list of mass changes (Phase I and Phase II reactions), background correction (unexpected transformations, des-alkylations)

Fragmentation analysis

Hypothetical fragmentation, comparison parent vs metabolite fragment spectra assignment of score (matches vs mismatches) → structure proposal + scoring

Automatic report system

Customizable excel file format (.xlsx), Database format (.xml format), Markush structure drawing from metabolite structures, Metabolite schemes (.pdf)

Qualitative and quantitative results

Reality Check: P450 Phenotyping

Combined Quant/Qual workflow

Goal:

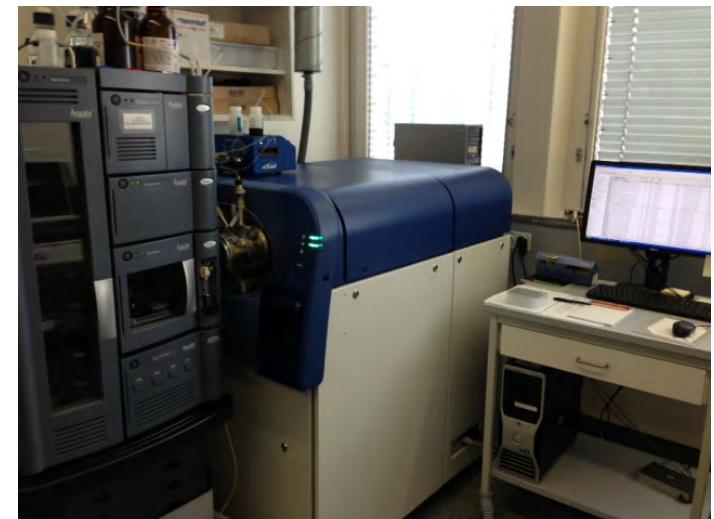
1. Identify P450 enzyme involved in drug clearance (**quant**)
2. Identify metabolites formed by individual enzymes (**qual & quant**)

Tools:

1. Matrix samples from 8 time point and 6 enzymes (48 samples)
2. AB Sciex 5600+ QqTOF mass spectrometer & fast gradient HPLC
3. IDA method based on intensity thresholds,
DBS active, no inclusion list
4. AB Sciex MetabolitePilot 1.5 software

Process:

1. Identify metabolites from individual samples
2. Matching across matrices & time points
3. Flexible reporting



Case Study: P450 Phenotyping

MetabolitePilot

Met ID process

1. Peak finding strategy: predicted metabolites (Phase I biotransformation list)
2. Generic parameters (threshold, mass tolerance, peak shape, ...)
3. Compound specific parameters (C-heteroatom bond cleavage, isotope pattern, product ion and neutral losses)
4. Formula prediction (include MS/MS)

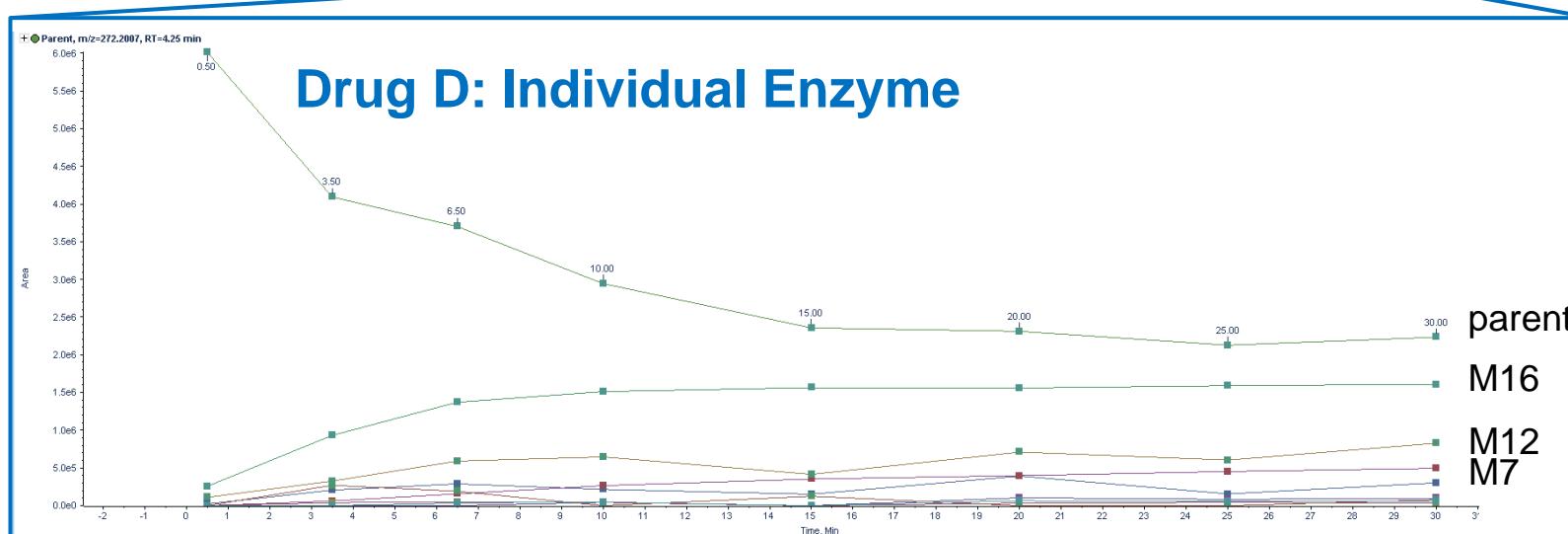
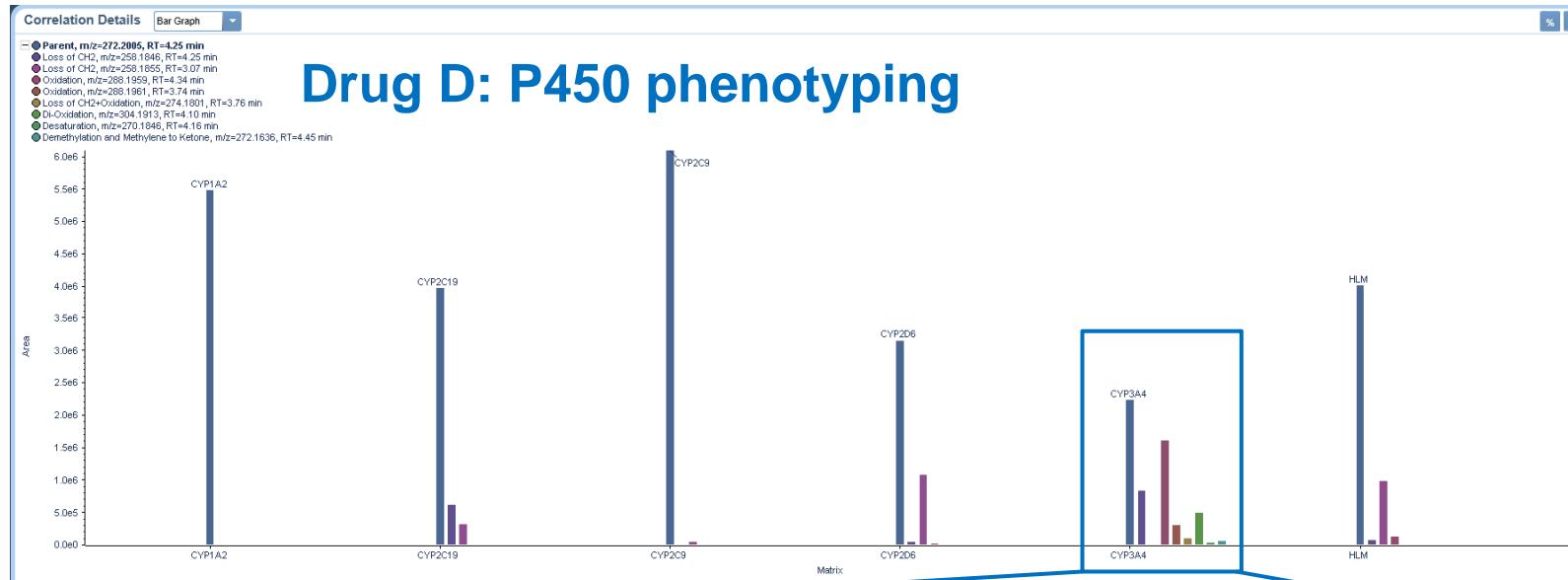
Results: CYP3A4 for drug D

Peak ID	Name	Formula	m/z	ppm	R.T. (min)	Peak Area	% Area	% Score
	Parent	C18H25NO	272.2005	-1.4	4.26	2.23E+06	35.0	93.8
M7	Di-Oxidation	C18H25NO ₃	304.1913	1.9	4.10	4.95E+05	7.8	81.9
M12	Loss of CH ₂	C17H ₂₃ NO	258.1846	-2.6	4.25	8.29E+05	13.0	95.4
M6	Loss of CH ₂ +Oxidation	C17H ₂₃ NO ₂	274.1801	-0.1	3.76	9.86E+04	1.5	88.4
M5	Oxidation	C18H ₂₅ NO ₂	288.1961	1.1	3.74	3.03E+05	4.7	75.0
M16	Oxidation	C18H₂₅NO₂	288.1959	0.5	4.34	1.61E+06	25.2	89.7
M8	Desaturation	C18H ₂₃ NO	270.1846	-2.2	4.16	3.45E+04	0.5	76.9
M17	Demethylation and Methylene to Ketone	C17H ₂₁ NO ₂	272.1636	-3.4	4.45	5.98E+04	0.9	87.4

- Excellent mass accuracy
- Very good match with manual data inspection
- Very few false negative results (threshold setting, MS/MS data was triggered)

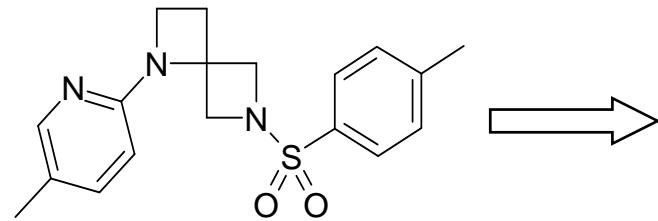
Case Study: P450 Phenotyping

MetabolitePilot, flexible reporting options



The Present & Future of Metabolite ID

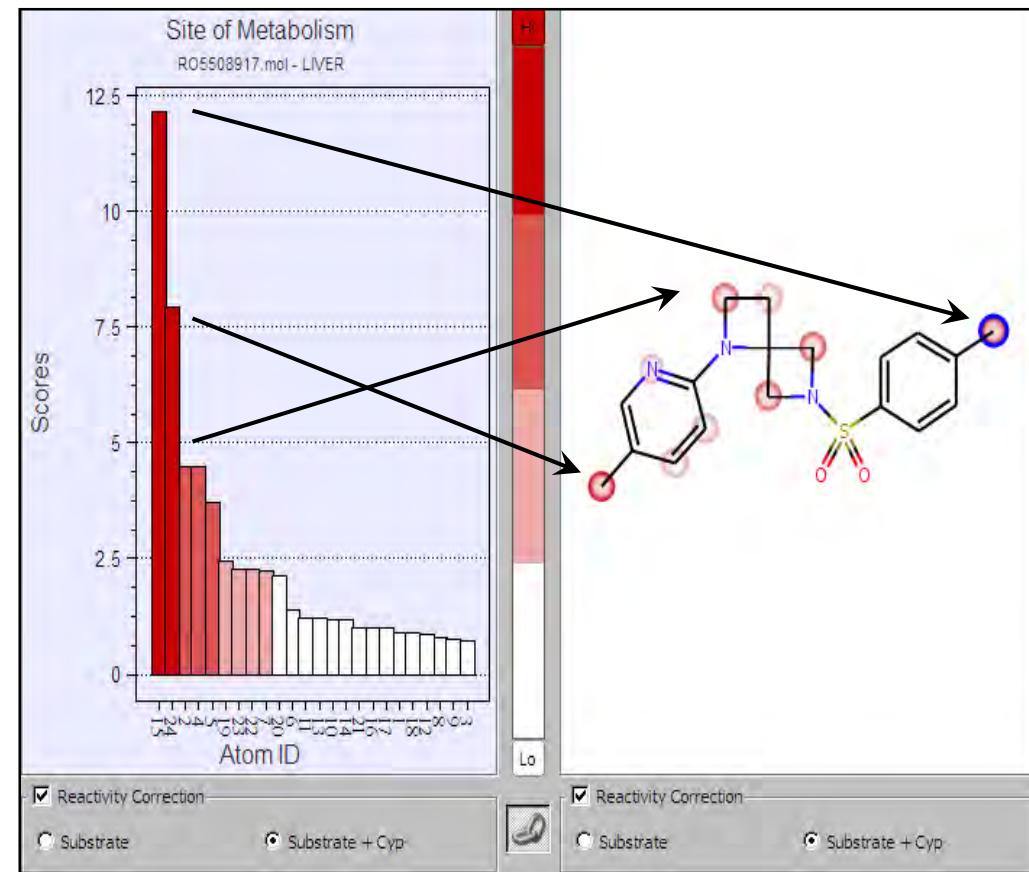
Computation chemistry approaches



In-silico prediction based on P450 binding site and chemical reactivity of drugs towards oxidation

SoM Prediction:
Generally good prediction of soft spot
70-80% success
(experiment within top 3)

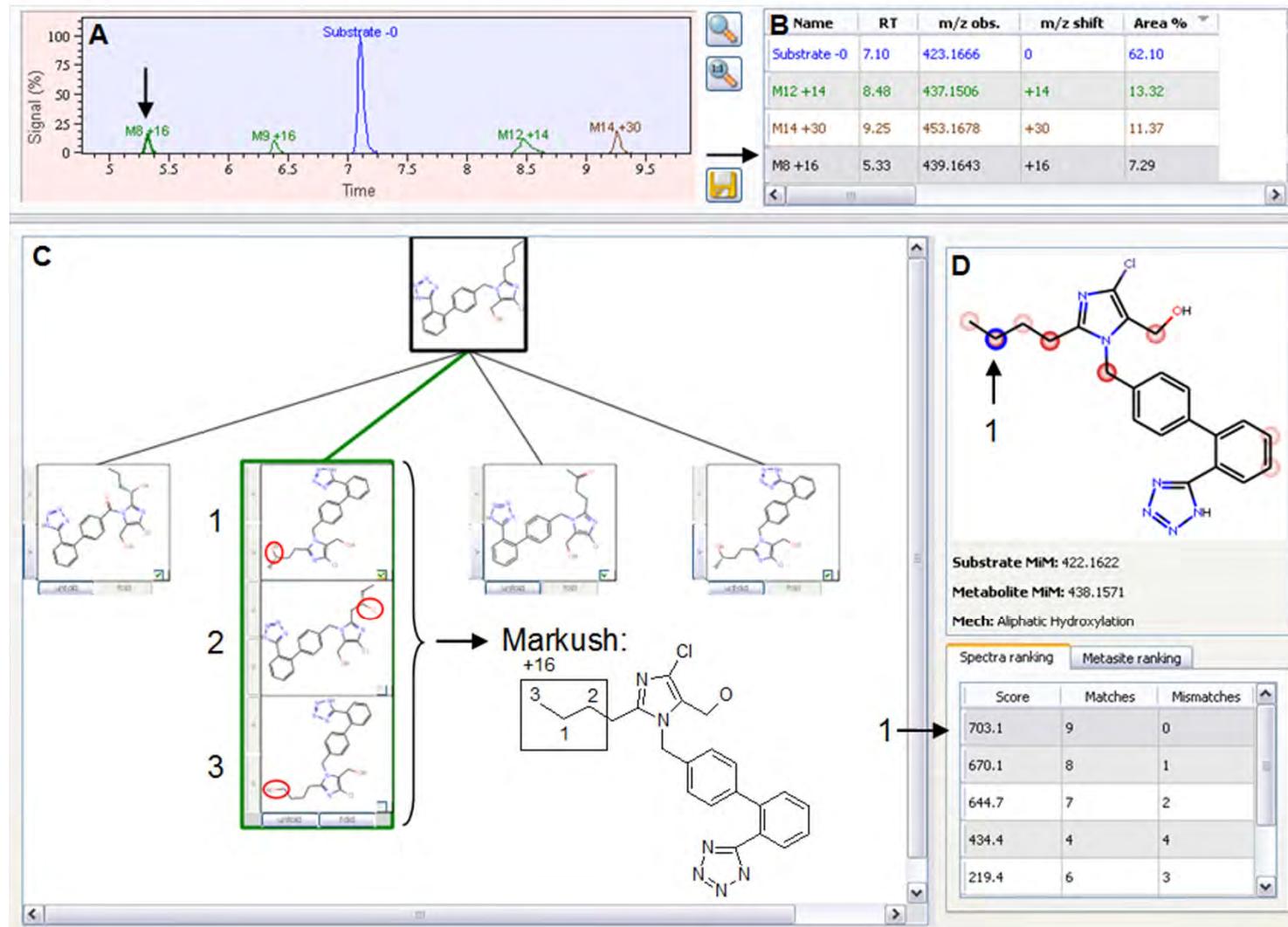
Used for prediction of likely SoM in conjunction with experimental MS/MS data and manual data interpretation



MetaSite (Molecular Discovery Ltd)

Making full Use of MS/MS Data

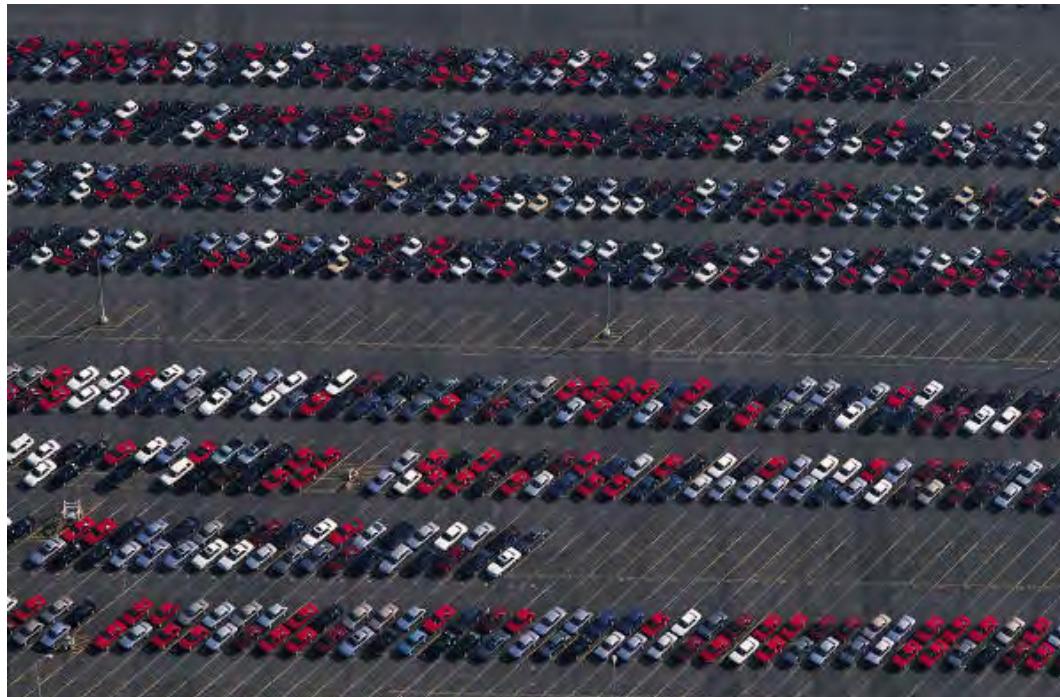
Blending computation chemistry and LC-MS/MS Data



Mass MetaSite (Lead Molecular Design)

Finally: What happens to all the data?

Diversity is nice, but....



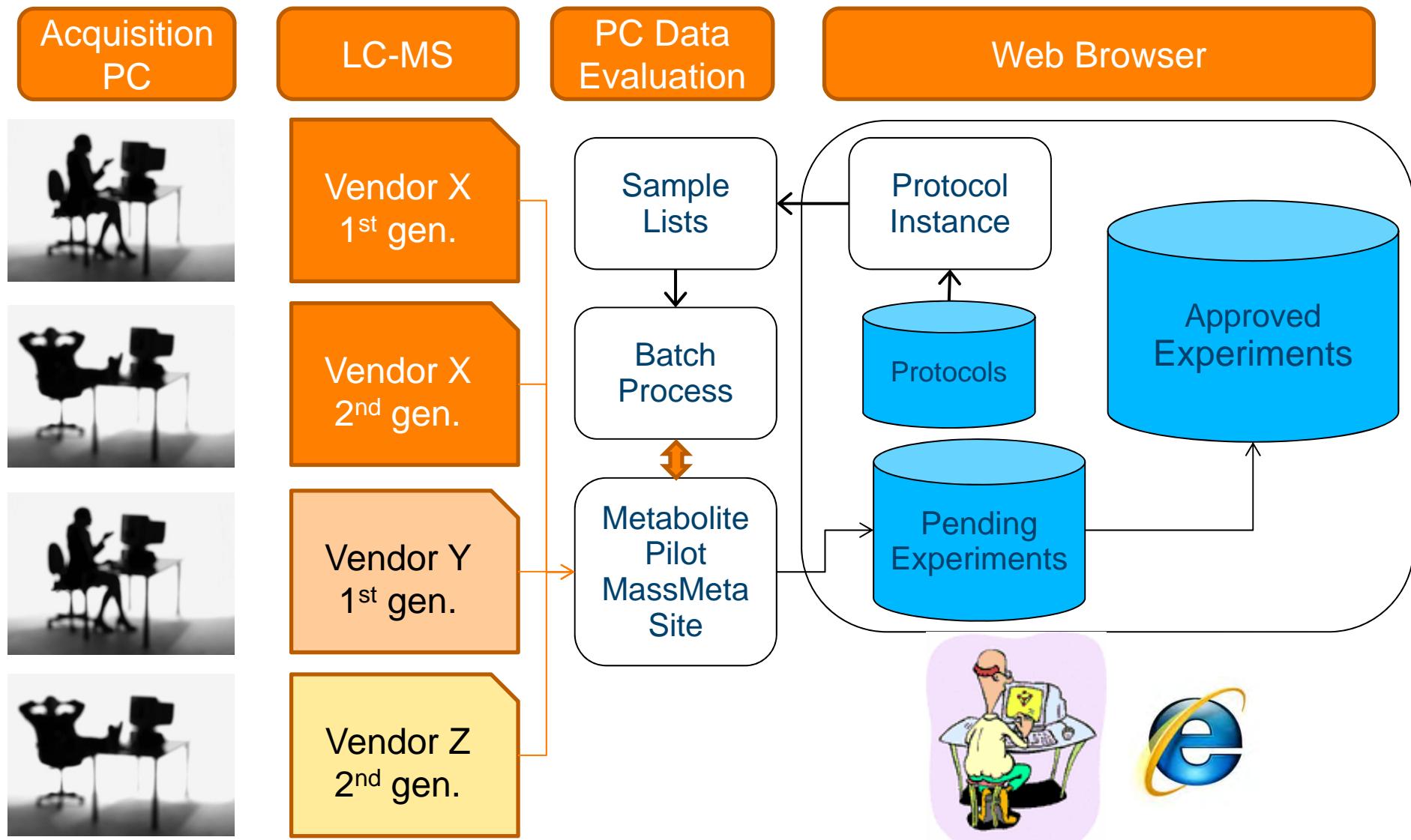
Some processes are “harmonized”.
But need to remember where I parked my car!

Traffic may become terrible...



Cross Platform DMPK Setup

Accommodating various workflows

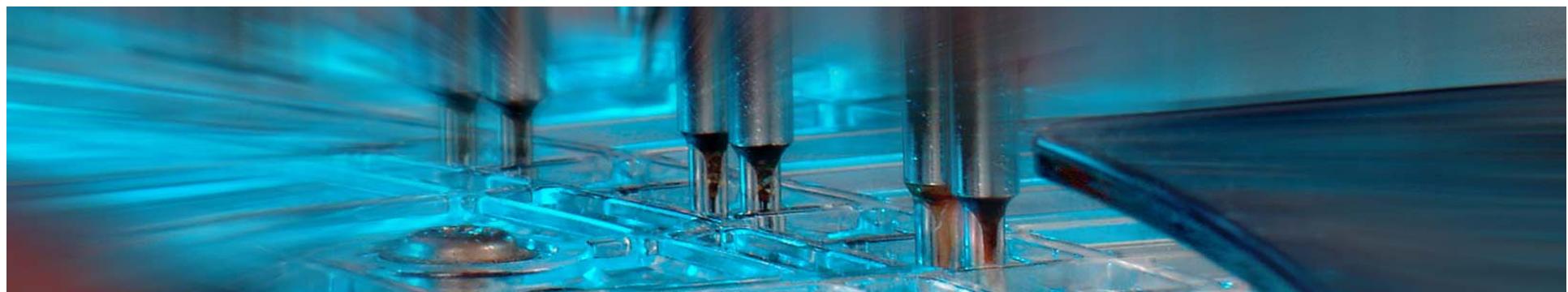


Conclusions & Outlook

- Emerging technologies (hardware & software) have facilitated workflows in Drug Discovery and Development
 - Unbiased acquisition and post-acquisition data processing
 - “All-in-one” approaches
 - Structure-based data analysis
- Qual/Quant workflows have become routine
 - Combined clearance / Met ID studies
 - Still need for “true” (not relative) quantitative results
 - Benefits of SRM acquisition (linearity, S/N)
 - SWATH & HRMS close to SRM performance
- Structure analysis
 - Computational approaches have emerged
 - Interrogation of LC-MS/MS data most promising

LC-MS/MS applications in biomarker research

Marieke Teppner
DMPK, Non Clinical Safety, pRED, F. Hoffmann-La Roche Ltd.



Content

- Introduction
- Experimental set-up
- Results: Method comparison between QTRAP 4000 and TTOF 5600+
- Biological application
- Conclusion

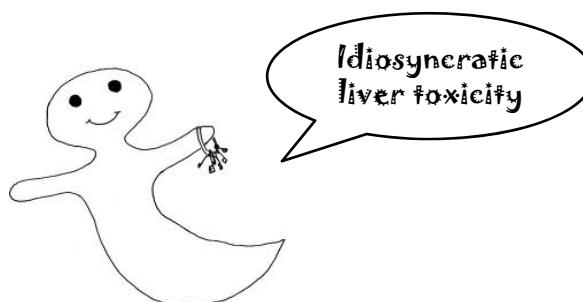
Introduction – Biomarker Research

- **Biomarker:**
Endogenous molecules utilized for objective diagnosis or monitoring of physiological, pathophysiological or pathological processes
 - Clinic: Observation or identification of disease state of patients
 - Pre-clinic: Testing of effect / side effect of drug candidates
- **Critical characteristics:**
 - Reliability (selectivity/sensitivity/robustness)
 - Mode of measurement (invasive vs noninvasive)

Introduction – Biomarker Research

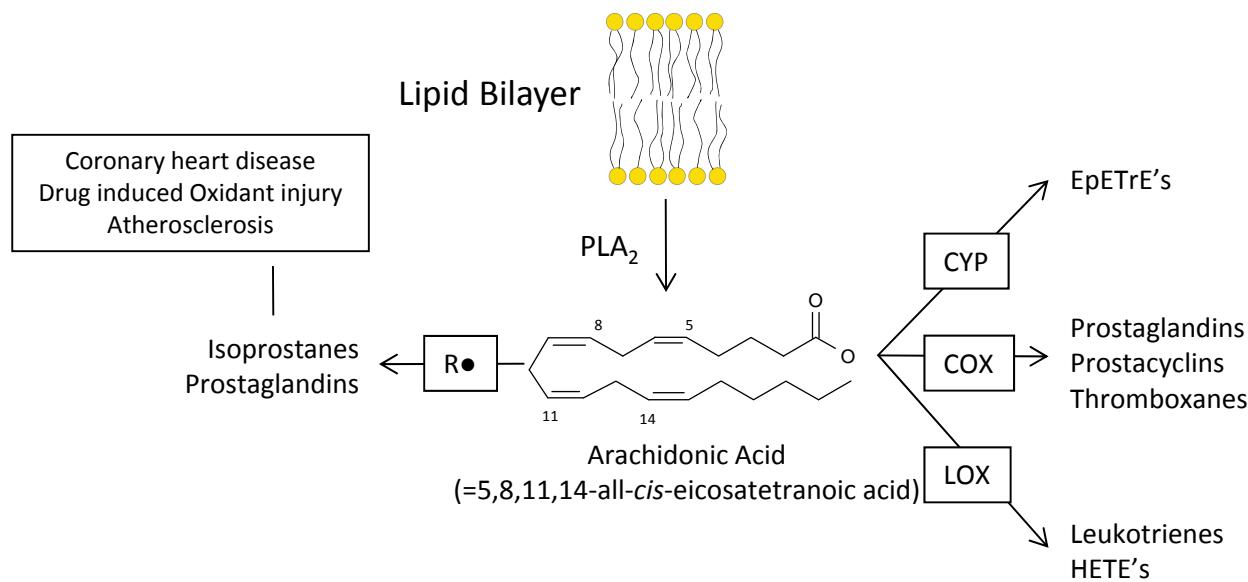
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 - Mode of measurement (invasive vs noninvasive)

Detection of severe
adverse effect as liver
toxicity (“DILI”)



Introduction – Isoprostanes as Biomarkers *in vivo* and *in vitro*

- Generation: Arachidonic acid metabolism



- Circulation in plasma, excretion to urine → potential target of investigation
- Generated in cells exposed to oxidative stress i.e. **hepatocytes** when cause is drug → suitable *in vitro* matrix

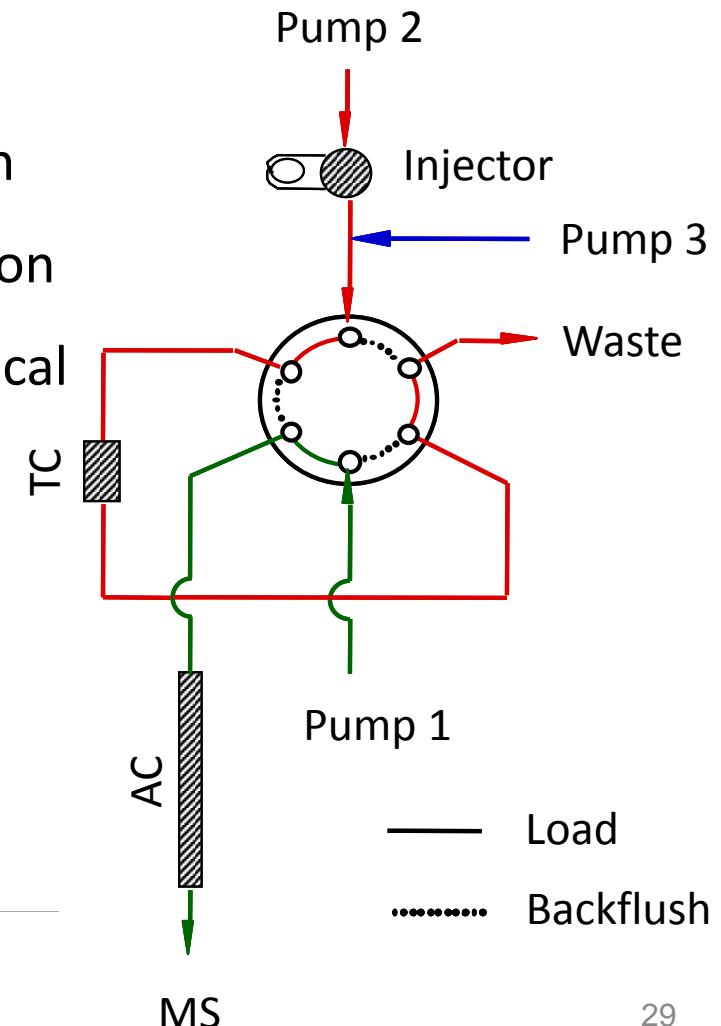
Experimental –

How to detect and quantify isoprostanes

1. Preparation step: Extraction of prostanoids from matrix
 - Solid phase extraction
 - liquid liquid extraction
 - immunoaffinity assays
2. Quantification step: Separation / Focusing of similar analytes + detection
 - LC-MS(/MS)
 - GC-MS
 - Immunoaffinity assays
 - Combination of 1 and 2 favorable to avoid work up steps
→ column switching LC set up for analyte enrichment

Experimental – Setup for integrated isoprostane measurement: Column switching LC

- Injection volume: 500 µL
- Run time 13.5 min, flow rate 0.4 mL/min
- Flush on trapping column + online dilution
- Backflush of retained analytes to analytical column + gradient elution
- MS/MS detection:
 - 4000 QTRAP (SRM)
 - 5600+ TTOF (FS + PIS → “HR-SRM”)



TC: YMC AQ, 20 x 2.1 mm, 5 µm, YMC Europe

AC: Atlantis T3, 100 x 2.1 mm, 1.8 µm, Waters

Experimental – MS options for QqQ versus TTOF instruments

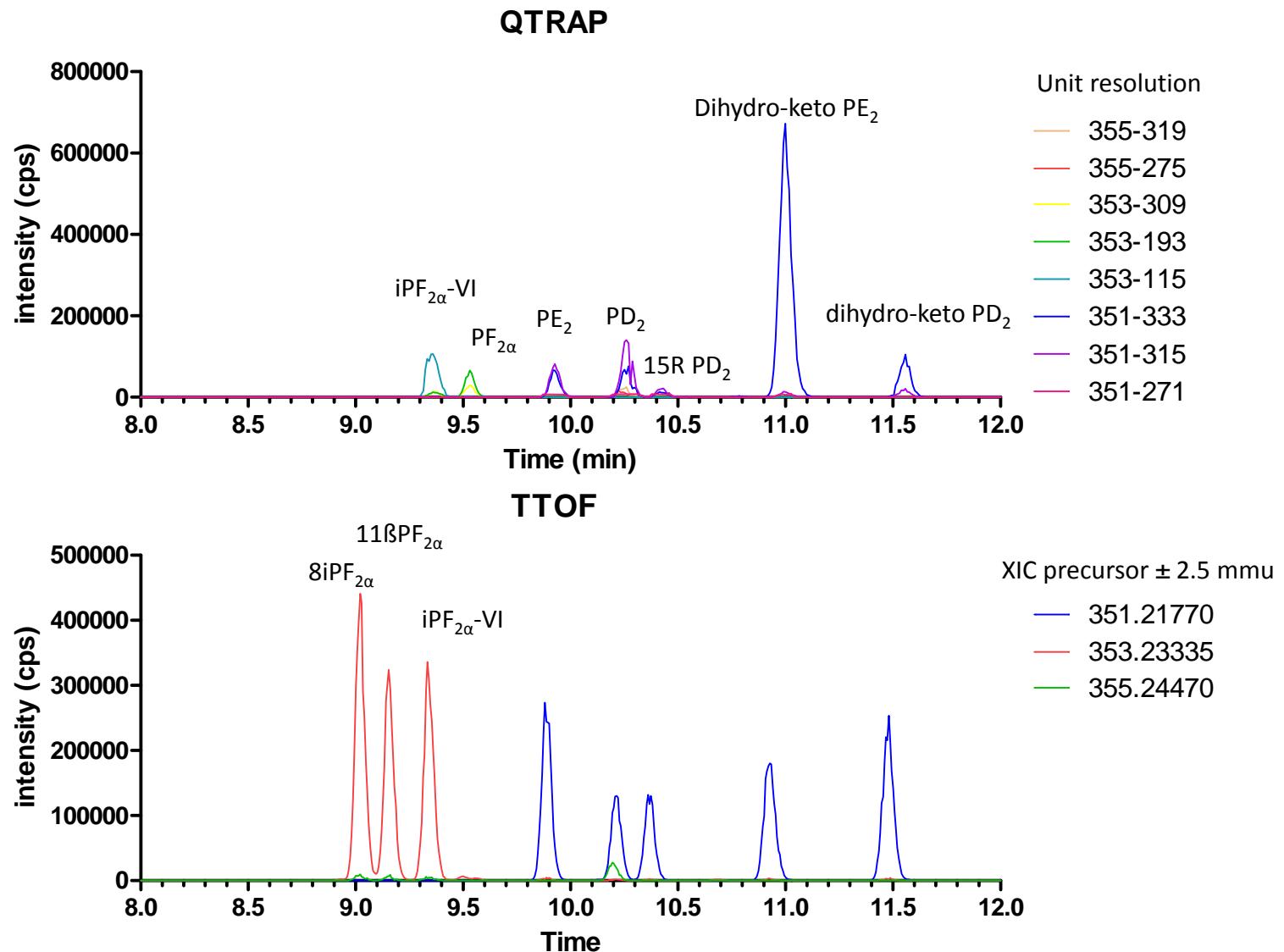
- Analytical requirements
 - 10 – 12 data points per peak
 - High selectivity
 - Wide linear range
- Available scan techniques
 - QqQ: Selected Reaction Monitoring (also: MRM)
best to exclude interferences and achieve linear calibration ranges of several orders of magnitude
 - TTOF:
 1. Full scan + data dependent scan
→ number of scans too small (many experiments)
 2. Full scan + product ion scan
→ allows to pick matching precursor + product ion

Optimization of Analytes

- QTRAP: Compound specific SRM transitions; TTOF: no optimization needed

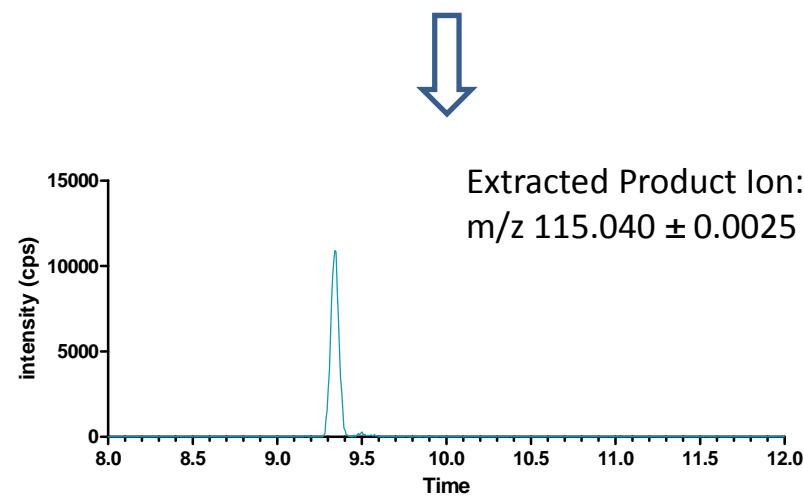
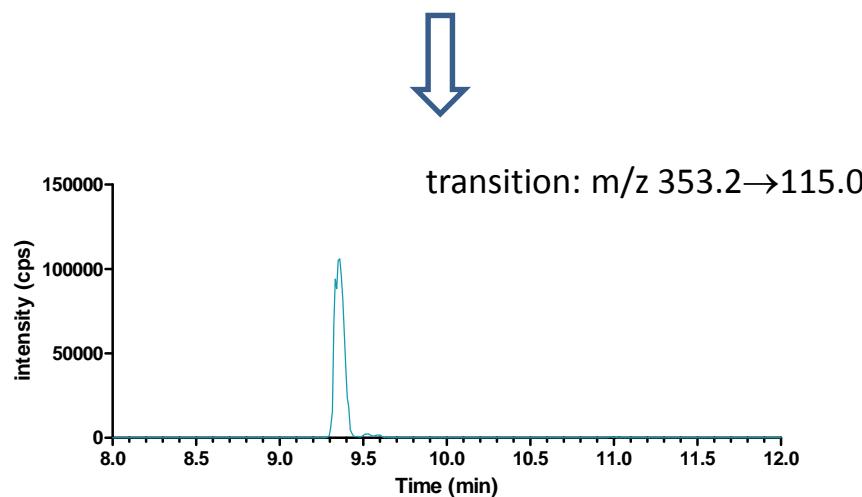
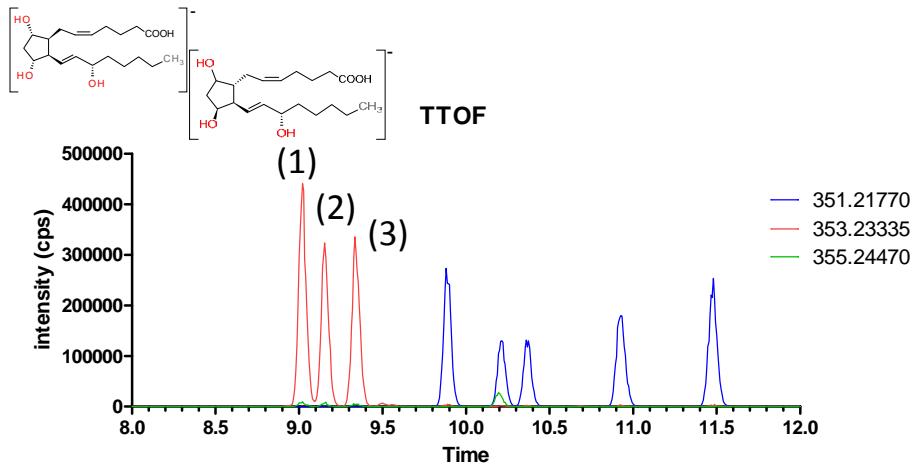
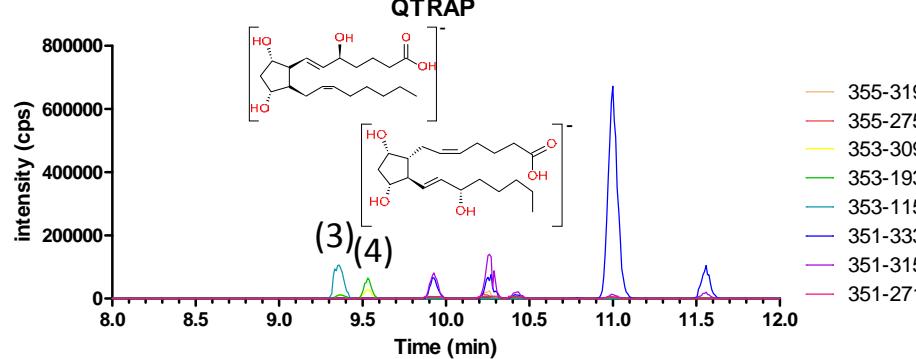
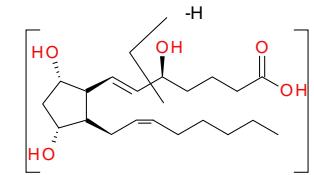
Short name (entry)	structure	precursor Q1 m/z	fragment Q3 m/z	DP (V)	CE (V)	CXP (V)
5-iso PF _{2α} -VI (1)		353.1	193.0	-65	-34	-13
		353.1	114.9	-80	-30	-9
PF _{2α} (2)		353.1	309.1	-60	-36	-15
		353.1	193.0	-65	-34	-13
PE ₂ (3)		351.1	315.0	-55	-18	-11
		351.1	271.1	-50	-25	-12
PD ₂ (4)		351.1	315.0	-55	-18	-11
		351.1	271.1	-50	-25	-12
15(R) PD ₂ (5)		351.1	315.0	-55	-18	-11
		351.1	271.1	-50	-25	-12
dihydro-keto PE ₂ (6)		351.1	333.0	-35	-16	-13
		351.1	315.0	-55	-18	-11
dihydro-keto PD ₂ (7)		351.1	333.0	-35	-16	-13
		351.1	315.0	-55	-18	-11
PD ₂ -d4 (8)		355.1	337.0	-65	-16	-17
		355.1	319.0	-55	-18	-11

Results – Overview reference spectra (5 ng/mL)



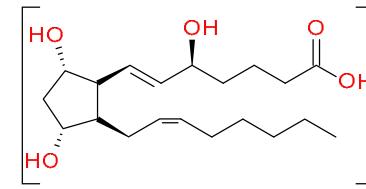
Results –

Overview reference spectra (5 ng/mL)



(1) – (4): Isobaric $F_{2\alpha}$ prostaglandins

Results – Comparison of Calibration data

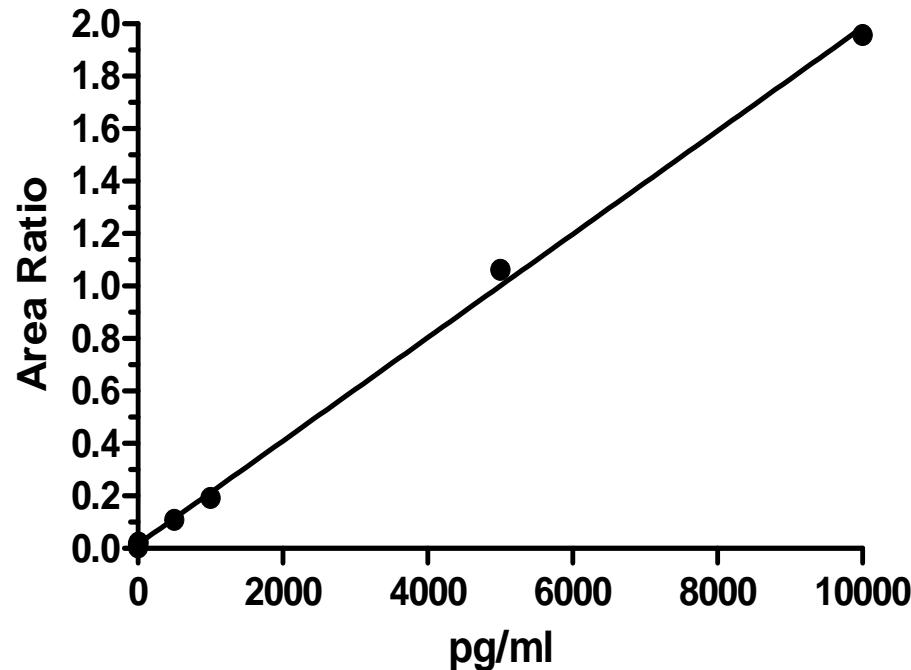
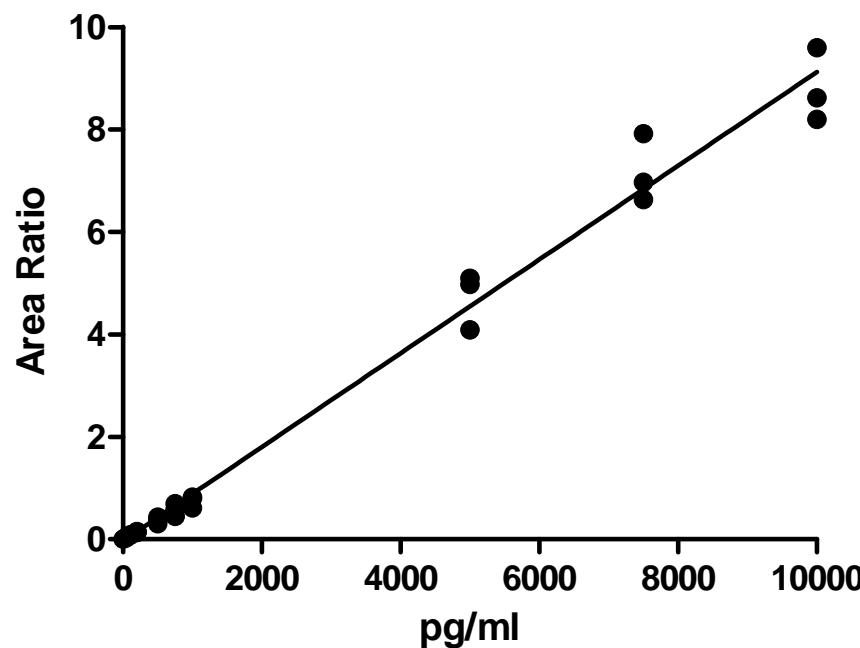


QTRAP: 5isoPF_{2α}-VI

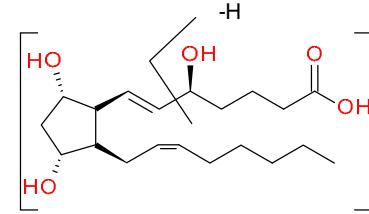
- Calibration from 0.02 to 10 ng/mL
- $y=0.0009152x-0.02752$
- $R^2=0.9892$

TTOF: 5isoPF_{2α}-VI

- Calibration from 0.005 to 10 ng/ml
- $y=0.0001971x-0.01568$
- $R^2=0.9985$

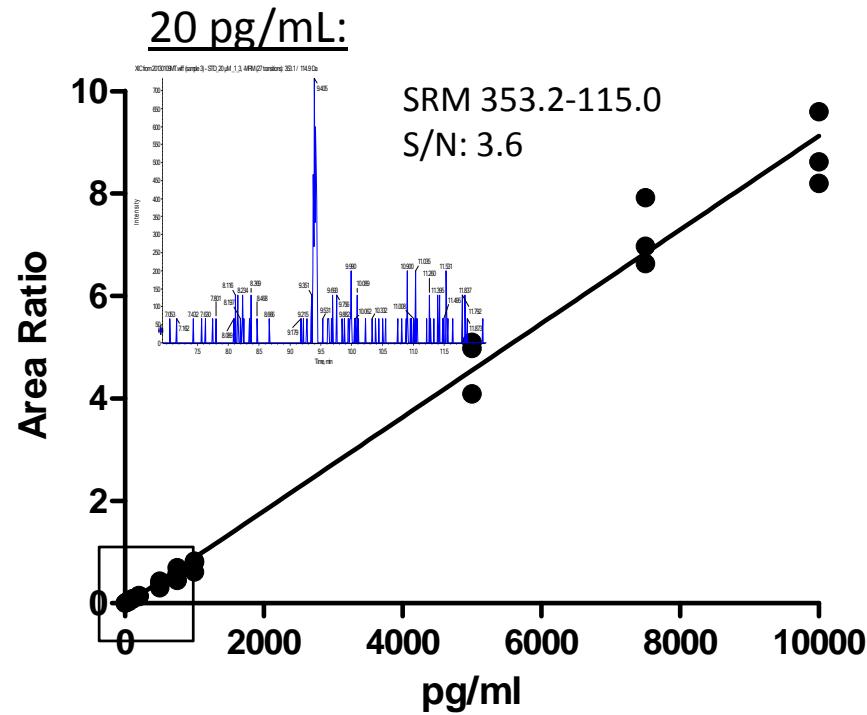


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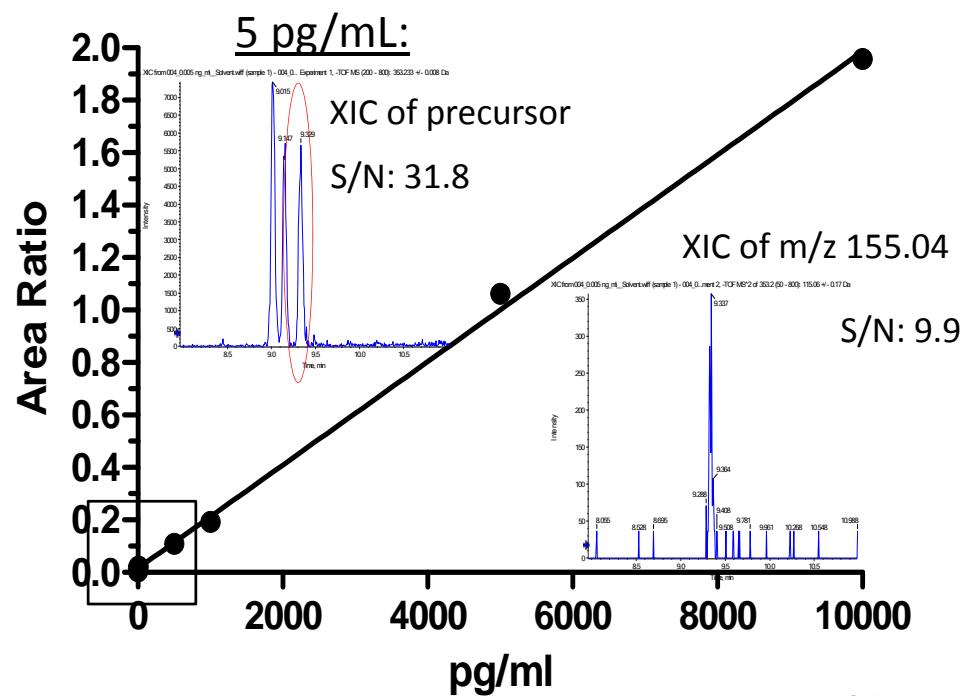
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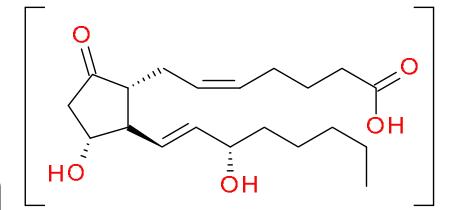


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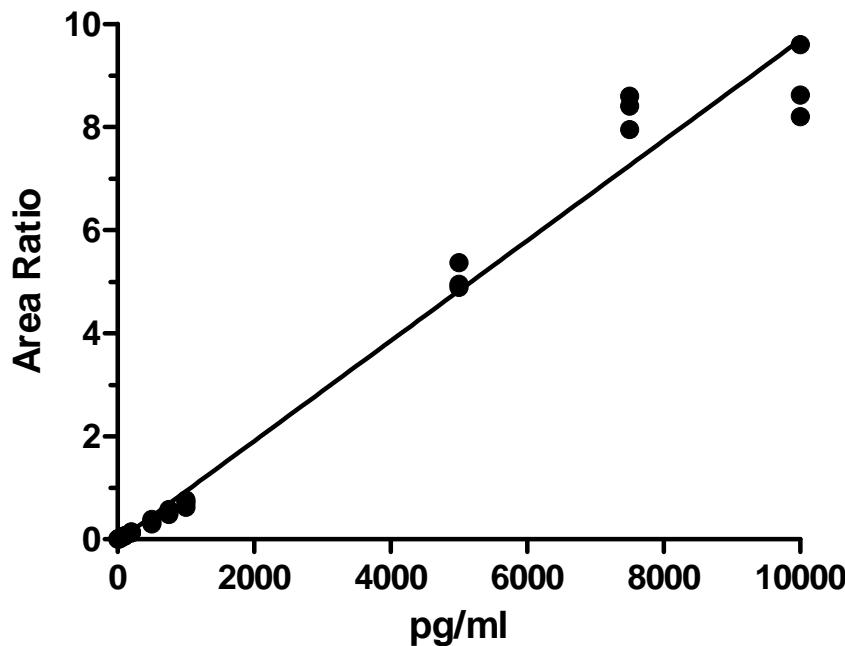


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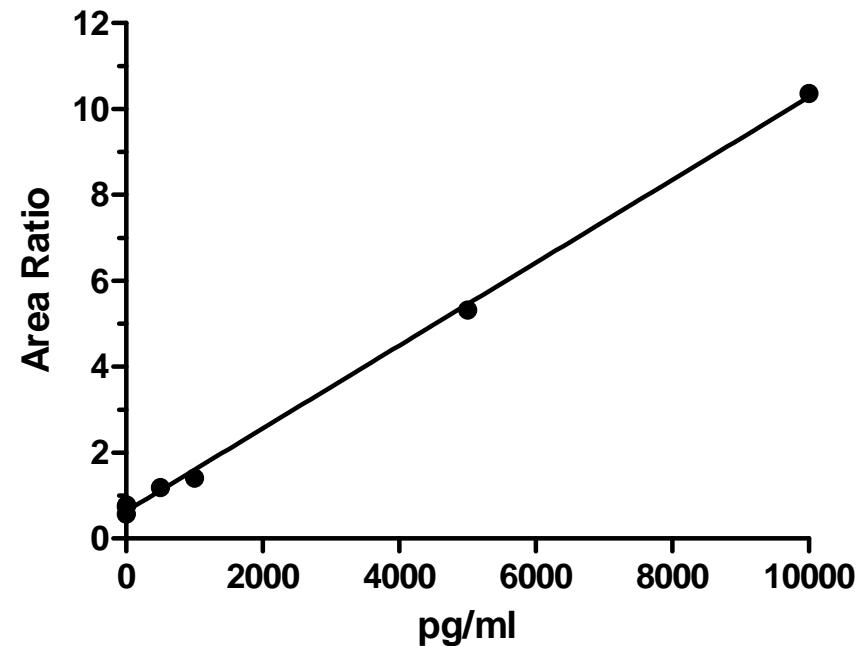
QTRAP: Prostaglandin D₂

- Calibration from 0.02 to 10 ng/mL
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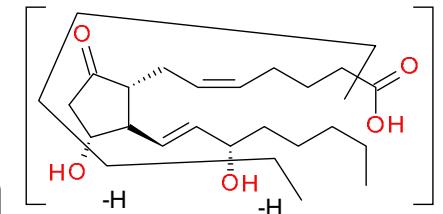


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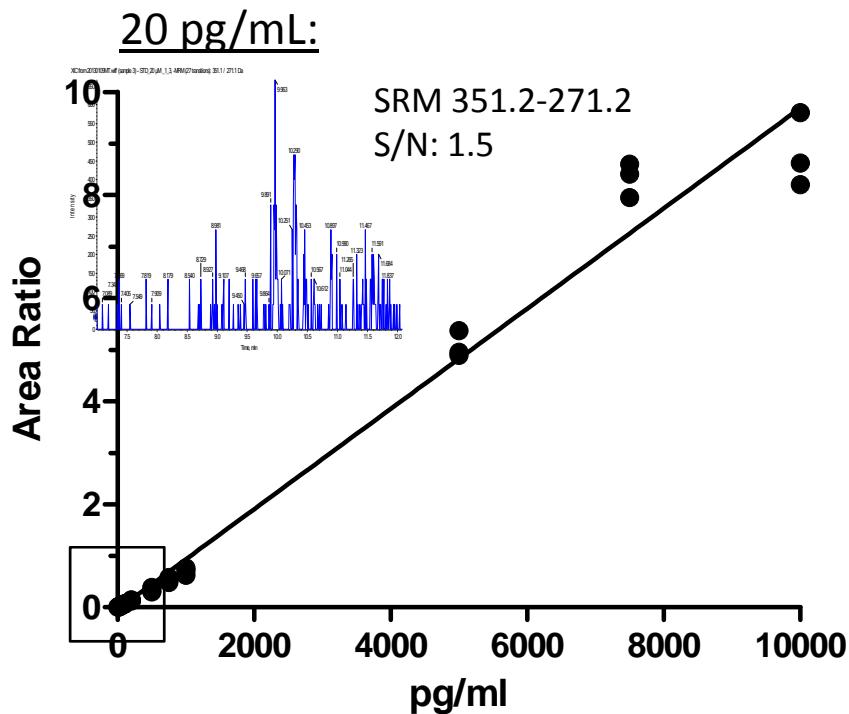


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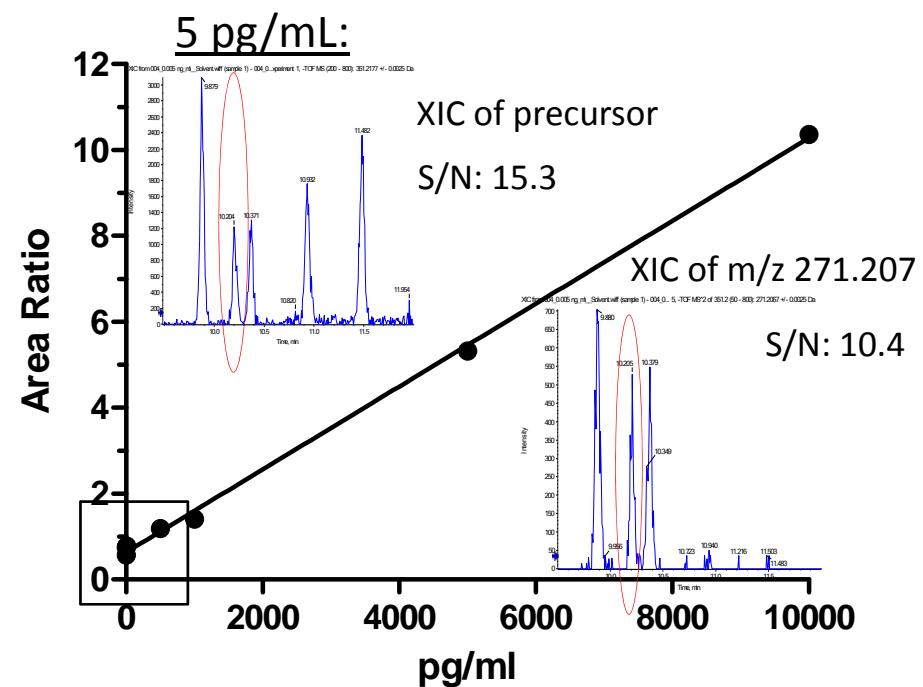
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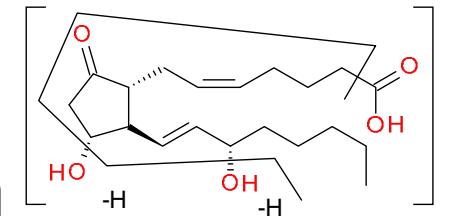


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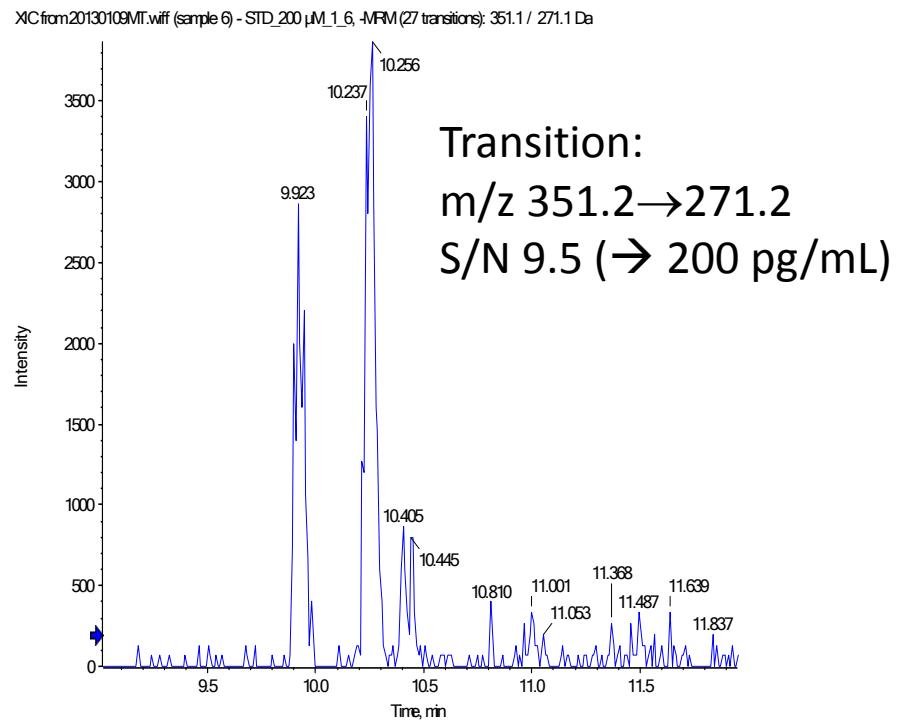


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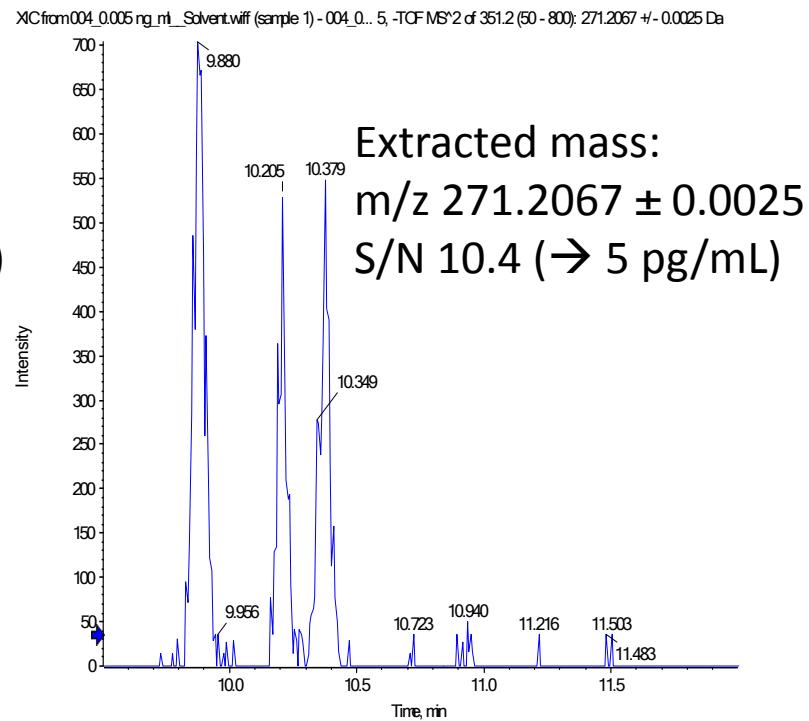
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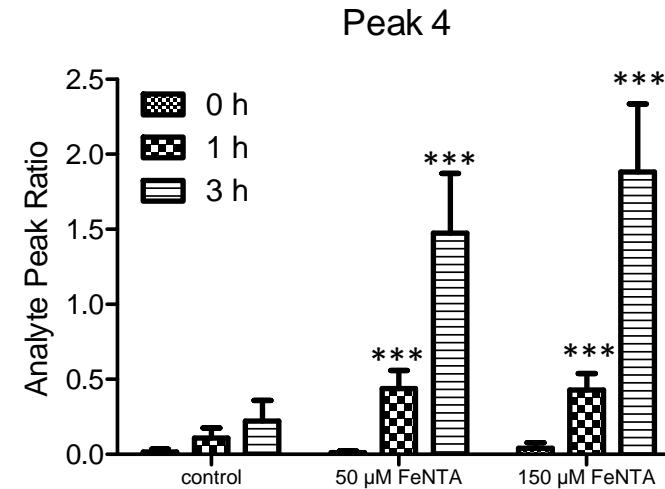
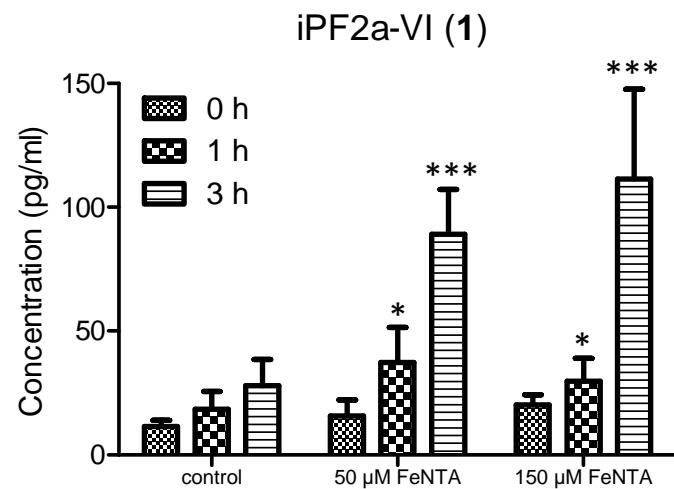
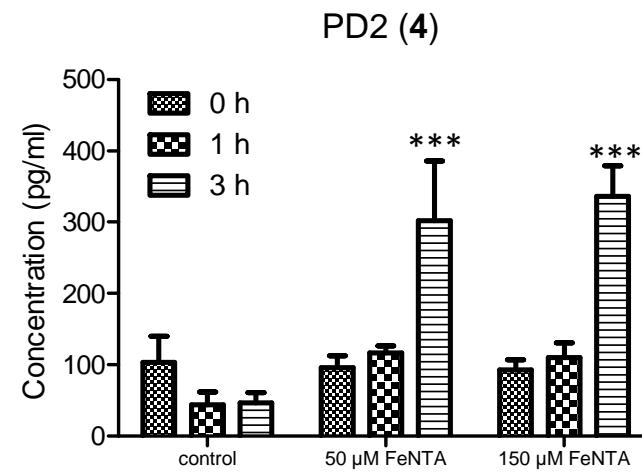
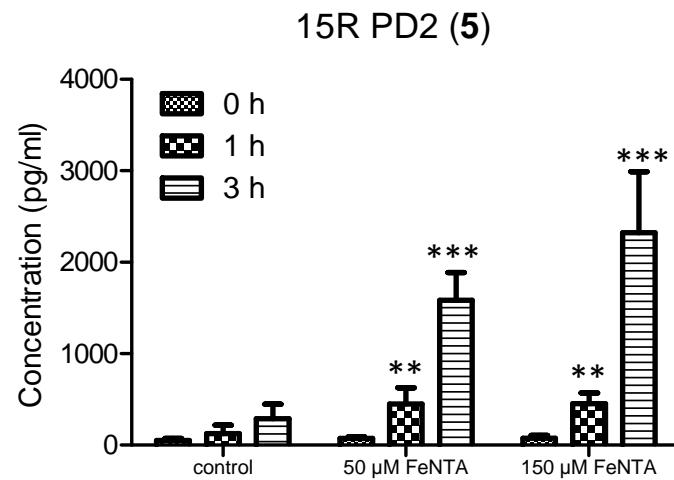


TTOF: Prostaglandin D₂

- Calibration from 0.005 to 10 ng/ml



Biological Application for Oxidative Stress Measurement: Rat hepatocytes



Conclusion

- The TripleTOF 5600+ can be used for quantitative biomarker analysis
- Most suited scan technique:
FullScan combined with ProductIonScan (“HR-MRM”)
 - Linear range: Comparable with triple quadrupole type instruments
 - Sensitivity gain in comparison with 4000 QTRAP for presented application
 - Selectivity: Achieved via accurate mass FullScan mode
Further enhanced by the use of specific product ions
 - Flexibility in post acquisition data evaluation
 - Define processing parameters
 - Most suitable product ion
 - Combined XIC of several selective product ions

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Thank you for listening!

Questions



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Marieke Teppner

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